

Union and Characterization of Molecularly Imprinted Polymer for Prasugrel Drug Base on 2-Hydroxyethyl Methacrylate Monomer in Biological and Pharmaceutical Samples

M. Ikuta¹, C. Matsumoto²

Department of Neonatology, Universitas Brawijaya, Jawa Timur 65145, Indonesia^{1,2}.



Abstract— In this examination work, a molecularly engraved polymer (MIP) was integrated for Prasugrel. In this technique, a polymer was first orchestrated utilizing 2-hydroxyethyl methacrylate (HEMA) as a monomer, N, N'- methylenebisacrylamide as a crosslinker, and azobisisobutyronitrile (AIBN) as an initiator. The fruitful blend of the MIP was affirmed by essential investigation (CHN), Fourier-change infrared (FT-IR) spectroscopy and checking electron microscopy (SEM). At that point, the impacts of different parameters, for example, pH, temperature, MIP limit, selectivity, adsorption energy and adsorption isotherms were examined by UV-Vis spectroscopy. The adjustment bend was plotted for Prasugrel in these conditions and direct range, point of confinement of recognition (LOD) and cutoff of measurement (LOQ) were accounted for. At long last, the presentation of the combined MIP as an adsorbent was considered under upgraded conditions for the extraction and assurance of Prasugrel in tablet framework and natural liquids, for example, pee and plasma by superior fluid chromatography (HPLC).

Keywords—Prasugrel, sub-atomic engraving polymer, natural liquids, pharmaceutical example.

1. Introduction

Prasugrel (synthetic equation: $C_{20}H_{20}FNO_3S$; sub-atomic weight: 273.442 g.mol⁻¹) is a white powder that effectively breaks up in methanol. It is a prodrug that must be changed over to a functioning metabolite by the liver, and is quickly changed into an inert thiolactone by hydrolases, and after that into a functioning metabolite by CYP catalyts. Prasugrel is bound to proteins (by around 98 percent) and discharged in the pee (68%) and dung (27%). It is endorsed for patients with hypertension or heart disappointment in which blood isn't siphoned adequately to the body. Physical and substance strategies, for example, superior fluid chromatography (HPLC) and UV spectrophotometry have been accounted for recognizing and estimating Prasugrel in natural liquids, and slim layer chromatography (TLC) for medication, plasma, and pee tests. Most medications are secluded and estimated utilizing HPLC in light of the fact that it appreciates high accuracy, extremity, affectability, and so on [1]- [6]. In this strategy, test planning is significant in light of the fact that it decreases the lattice impact in natural liquids and yields solid outcomes [7]. Molecularly engraved polymers (MIPs) are another gathering of manufactured materials with explicit acknowledgment locales. MIPs are created as a 3-D polymeric system with solid crosslinkers in the copolymerization response between a utilitarian monomer and a crosslinker within the sight of a format atom [8], [9]. After the MIPs are delivered, the format atoms are expelled from the acquired structure because of which restricting locales are made with the structure, size, and utilitarian gatherings coordinating those of the layout. Different techniques are utilized to create MIPs. Fluid polymerization has been utilized in many reports on setting up these polymers since it is a less complex technique. The best outcomes in incorporating MIPs are gotten by utilizing little formats; subsequently, Prasugrel was utilized as the layout in the present research. Monomer determination requires incredible consideration with the goal that it can make dynamic and specific locales for the format. We utilized 2-hydroxyethyl methacrylate as the monomer in light of the

fact that the carboxyl gathering in its structure empowers it to pull back or give hydrogen particles because of pH changes in the earth and, in this manner, can be viewed as pH touchy.

We expected to deliver an exceptionally particular and pH delicate polymer, and consolidate MIP-based strong stage extraction (SPE) and HPLC to identify and quantify Prasugrel in organic liquids and medication tests.

2. EXPERIMENT

2.1 Instruments

IR spectra were recorded on an IR Fourier change spectrometer (FT-IR-4100, made by Jasco Inc., Easton, Maryland, USA), warm gravimetric examination (TGA) was performed utilizing a warm analyzer (made by the Shimadzu Corp. in Japan), an examining electron magnifying lens or SEM (Philips XL-30 FEG-SEM) was utilized for SEM investigation, basic examination by a CHN analyzer (made by the Lineis Messgerate GmbH Company in Hanau, Germany, Vario El), and UV/Vis spectroscopy by an UV/Vis spectrophotometer (V-530, created by the Jasco Inc., Easton, Maryland, USA).

2.2 Reagents and Solutions

2-hydroxyethyl meth (HEMA) and N, N' - methylenebisacrylamide, hexadecane, acetonitrile, methanol, dipotassium phosphate, and phosphoric corrosive were purchased from the Merck Company (Darmstadt, Germany), azobisisobutyronitrile (AIBN) from the Across Company (New Jersey, USA), and sodium dodecyl sulfate (SDS) from the Fluka Chemical Corp. (Buchs, Switzerland).

All reagents were HPLC grade. The stock arrangement (100 mg L^{-1}) was set up by dissolving 5 mg of Prasugrel in 50 ml of methanol, and the pH of the arrangement was balanced utilizing 0.01M NaOH or 0.01M hydrochloric corrosive.

3. Conclusion

In this investigation, rice, one of the principle substance by and large waste, was changed over into combustible gas and carbonaceous adsorbent by pyrolysis with sodium hydroxide. Crude rice can be changed over into a combustible gas, chiefly including hydrogen, methane, carbon monoxides and carbon dioxide gases. With expanding the expansion of sodium hydroxide or warming temperature, the combustible gas including higher substance of hydrogen and methane can be acquired. The heaviness of the buildup after pyrolysis with proportional sum expansion of sodium hydroxide at $700 \text{ }^\circ\text{C}$ is 1 % of crude rice test, and the buildup had a permeable structure with high explicit surface territory, which is proportionate to industrially accessible initiated carbon.

4. References

[1] W. L. Baker and C. M. White, "Role of Prasugrel, a novel P2Y₁₂ receptor antagonist, in the management of acute coronary syndromes," American Journal of Cardiovascular Drugs, vol. 9, no. 4, pp. 213-229, 2009.

- [2] S. D. Wiviott, E. Braunwald, and C. H. McCabe, "Prasugrel versus Clopidogrel in patients with acute coronary syndromes," *N. Engl. J. Med.*, vol. 357, no. 20, pp. 2001–2015, 2007.
- [3] Food and Drug Administration (United States), FDA Announces New Boxed Warning on Plavix: Alerts Patients, Health Care Professionals to Potential for Reduced Effectiveness, Press Release, March 13, 2010.
- [4] FDA Drug Safety Communication: Reduced Effectiveness of Plavix (clopidogrel) in Patients Who Are Poor Metabolizers of the Drug, Drug Safety and Availability, Food and Drug Administration (United States), March 12, 2010.
- [5] D. J. Angiolillo, J. F. Saucedo, and R. DeRaad, "Increased platelet inhibition after switching from maintenance Clopidogrel to prasugrel in patients with acute coronary syndromes," *J. Am. Coll. Cardiol.*, vol. 56, pp. 1017-1023, 2010.
- [6] M. O'Riordan. (2011). Switching from clopidogrel to prasugrel further reduces platelet function. [Online]. Available: <http://www.theheart.org>
- [7] V. Pichon and F. Chapuis-Hugon, "Role of molecularly imprinted polymers for selective determination of environmental pollutants—A review," *Anal. Chim. Acta*, vol. 622, no. 1-2, pp. 48-61, 2008.
- [8] D. Wang, S. Pyo Hong, G. Yang, and K. Ho, "Caffeine molecular imprinted microgel spheres by precipitation polymerization," *Korean J. Chem. Eng.*, vol. 20, no. 6, pp. 1073-1076, 2003.
- [9] I. Dakova, I. Karadjova, V. Georgieva, and G. Georgiev, "Ion-imprinted polymethacrylic microbeads as new sorbent for preconcentration and speciation of mercury," *Talanta*, vol. 78, no. 2, pp. 523-529, 2009.
- [10] H. A. Panahi, J. Morshedian, N. Mehmandost, E. Moniri, and I. Y. Galaev, "Grafting of poly [1-(N, N-bis-carboxymethyl) amino-3-allylglycerol-co-dimethylacrylamide] copolymer onto siliceous support for preconcentration and determination of lead (II) in human plasma and environmental samples," *J. Chromatogr. A.*, vol. 1217, no. 32, pp. 5165-5172, 2010.
- [11] R. K. Rajendiran, V. K. Sekar, B. D. Namadevan, J. K. Annamalai, and S. Devarajan, "UV-spectrophotometric and RP-HPLC methods for the estimation of prasugrel hydrochloride in bulk and tablet formulation," *Int. J. Pharm. Pharm. Sci.*, vol. 6, no. 1, pp. 220-225, 2014.
- [12] S. D. Darshali, S. B. Bharati, and G. W. Sanjay, "Stability indicating method for quantitation of prasugrel hydrochloride in presence of its degradation products," *International Journal of Pharmacy & Technology*, vol. 4, no. 3, pp. 4711-4720, 2012.
- [13] R. W. Enaksha, T. Ye, J. R. Kenneth, M. V. Elizabeth, J. W. Govinda, K. Atsushi, A. F. Nagy, "Stereo selective metabolism of prasugrel in humans using a novel chiral liquid chromatography-tandem mass spectrometry method," *Drug Metabolism and Disposition*, vol. 35, no. 6, pp. 917-921, 2007.
- [14] T. C. Borole, R. Mehendre, M. C. Damle, and K. G. Bothara, "Development and validation of stability indicating HPTLC method for determination of Prasugrel," *J. Chem. Pharm. Res.*, vol. 2, no. 4, pp. 907-913, 2010.

- [15] I. Srikanth, P. Sharmila, K. Vijayabharathi, M. Raju, M. LakshmaNaik, and K. Nagarjuna, "A validated reverse phase HPLC method for the estimation of prasugrel hydrochloride in pharmaceutical dosage forms," *JITPS*, vol. 2, no. 5, pp. 140-148, 2011.
- [16] C. Baggiani, C. Giovannoli, L. Anfossi, and C. Tozzi, "Molecularly imprinted solid-phase extraction sorbent for the clean-up of chlorinated phenoxyacids from aqueous samples," *J. Chromatogr. A*, vol. 938, no.1-2, pp. 35-44, 2001.
- [17] G. Theodoridis, A. Kantifes, P. Manesiotis, N. Raikos, and T. P. Heleni, "Preparation of a molecularly imprinted polymer for the solid-phase extraction of scopolamine with hyoscyamine as a dummy template molecule," *J. Chromatogr. A*, vol. 987, no.1-2, pp. 103-109, 2003.
- [18] E. P. C. Lai and S. G. Wu, "Molecularly imprinted solid phase extraction for rapid screening of cephalexin in human plasma and serum," *Anal. Chim. Acta*, vol. 481, no. 2, pp. 165-174, 2003.
- [19] L. Langmuir, *J. Am. Chem. Soc.*, vol. 40, p. 1361, 1918.
- [20] H. M. A. Freundlich, "Over the adsorption in solution," *J. Phys. Chem.*, vol. 57, pp. 385-471, 1906.
- [21] M. I. Tempkin and V. Pyzhev, "Kinetics of ammonia synthesis on promoted iron catalyst," *Acta Phys Chim USSR*, vol. 12, pp. 327-356, 1940.
- [22] G. Jin and Y. Tang, "Evaluation of a novel silica-supported sol-gel sorbent prepared by a surface molecular imprinting technique for the selective separation of estazolam from human plasma," *Microchim. Acta*, vol. 165, p. 143, 2009.
- [23] H. Guo and X. He, "Study of the binding characteristics of molecular imprinted polymer selective for cefalexin in aqueous media," *Fresenius J. Anal. Chem.*, vol. 368, pp. 461-465, 2000.
- [24] D. M. Han, G. Z. Fang, and X. P. Yan, "Preparation and evaluation of a molecularly imprinted sol-gel material for on-line solid-phase extraction coupled with high performance liquid chromatography for the determination of trace pentachlorophenol in water samples," *J. Chromatogr. A*, vol. 1100, no. 2, pp. 131-136, 2005.
- [25] X. Jiang, C. Zhao, N. Jiang, H. Zhang, and M. Liu, "Selective solid-phase extraction using molecular imprinted polymer for the analysis of diethylstilbestrol," *Food Chem.*, vol. 108, no. 3, pp. 1061-1067, 2008.



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